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be a promising cancer therapy. Recently, we have synthesized new HDAC inhibitors and among more than 100 lead compounds, SK-7068 was found to be the most effective in various cancer cells. The growth inhibition effect by SK-7068 was 4 times potent than that by MS-275, a benzamide-based HDAC inhibitor. As expected for a HDAC inhibitor, SK-7068 inhibited cellular HDAC enzymatic activity by 90% and induced the acetylation of H3 and H4. Although more than 9 isotypes of HDAC have been identified so far, it is unclear whether HDAC inhibitors specifically target specific isotypes of HDACs. To address this, we have tested whether the level of HDAC expression is correlated with sensitivity to SK-7068. SK-sensitive cells showed relatively higher level of HDAC1, suggesting that SK-7068 might selectively target HDAC1 isotype. SK-7068 treatment induced cell cycle arrest at G1-phase and also activated mitochondrial-caspase dependent apoptosis. Interestingly, SK-7068 treated cells were arrested at M-phase as well, which was demonstrated by increased mitotic index and dense chromatin formation. This is the first observation that a HDAC inhibitor might directly affect the mitotic check point regulation. Taken together, a novel HDAC inhibitor SK-7068 shows diverse and unique cellular effects in contrast with other HDAC inhibitors. Finally, cDNA microarray analysis revealed that transcriptional silencing of some novel genes was reversed after treatment with SK-7068. The experiments to understand the biologic significance of these target genes are now underway and final result will be presented in the meeting.

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Linking genotype to phenotype: production and large scale functional analysis of gene knockouts in human cancer cells

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To fully realize the impact of genomics technologies such as expression profiling, proteomics, metabolic pathway modeling, or SNP analysis, sequence information from the human genome database needs to be directly tied to drug discovery. Since gene inactivation (knockout) is predictive for effects of drug candidates, genetic methods to inactivate human genes are key to linking genotype with phenotype and for discovering the therapeutic utility of novel and known genes. Here we describe a gene knockout technology designed to directly link genotype with phenotype on a large scale. We use a plasmid-based vector to gene-trap and recover genes expressed in human cancer cells. From these, a library of over 7,500 knockout vectors has been generated. Homologous recombination with these vectors is exceptionally high (up to 10-1) allowing for creation of arrays of cells with gene knockouts. The array of cells (either as single copy or multiple copy knockouts) can be subjected to multiple assays to determine which genes are essential for genesis or maintenance of various attributes of cancer cells or to determine which genes, when inactivated, sensitize cells to various chemotherapeutic agents and radiation. The gene-trap vectors contained secretory alkaline phosphatase (SEAP) that allowed for monitoring of endogenous trapped promoter activities. We created a clonal SEAP-reporter library in the pancreatic tumor cell line MiaPaCa-2, and screened for genes responsive to the activation of the erb-B-receptor family; erb-B1, erb-B2 and erb-B3 but not erb-B4 were found to be expressed in MiaPaCa-2 cells. Stimulation of the MiaPaCa-2 library with EGF, TGF-alpha, Amphiregulin, Betacellulin or Heregulin-alpha resulted in increased SEAP activity in 3% of the clones. 75% of these clones responded only to TGF-alpha, 10% were responsive to EGF, Amphiregulin and TGF-alpha, and 15% responded to both EGF and TGF-alpha. Further screening of the MiaPaCa-2 library for increased sensitivity to gemcitabine, 5-FU, cisplatin, camptothecin, and DNA damaging radiation yielded a number of clones representing potential drug targets. The described technologies allow us to establish a direct link between genes represented in the human genome database and drug target discovery. Furthermore, the ability to create the genetic lesion in alternative cancer cells using highly efficient homologous recombination vectors allows us to evaluate these genes in diverse disease backgrounds.

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Differential kinetic properties of monomeric and oligomeric phosphorylated c-met

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One mechanism for regulating receptor tyrosine kinases (RTKs) is for the active and inactive states of the enzyme to have different kinetic properties. An important aspect of this regulatory mechanism is how oligomerization-

induced conformational changes modulate the kinetic properties of RTKs. Recombinant TPR-MET protein, a functionally active translocation oncoprotein derivative of c-MET, has been expressed and purified for enzymatic analysis. This naturally occurring oncoprotein contains the cytoplasmic domain of the cMET receptor fused to a coiled coil motif from the nuclear pore complex (TPR). CytoMET, a monomeric analog of TPR-MET, has also been expressed and purified for differential enzymatic analysis. ATP and peptide substrates corresponding to in vivo phosphorylation sites within the cytoplasmic domain of the receptor have been kinetically characterized for fully phosphorylated TPR-MET and fully phosphorylated CytoMET. Our kinetic data shows that the catalytic activity of TPR-MET is modulated by oligomerization, independent of activation loop phosphorylation, and thus suggests that oligomerization-induced conformational changes occur within the cytoplasmic domain of RTKs. These results have significant implications for structure-based design of RTK inhibitors and the development of a detailed mechanistic model of BTK activation

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Correlation between NF-kB activity and lkB degradation in tumor cells using bioluminescent reporters

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Recent evidence indicates that NF-kB and the signaling pathways that are involved in its activation play an important role in tumor development. Constitutively activated NF-kB transcription factors have been associated with tumorigenesis, cancer cell proliferation, preventing apoptosis and increasing the angiogenic and metastatic potential of a tumor. NF-kB is regulated in part by its association with its inhibitor, IkB. IkB kinase phosphorylates IkB in response to agents that activate NF-kB. Phosphorylated IkB is ubiquitinated and rapidly degraded, releasing NF-kB and allowing it to translocate to the nucleus and activate transcription of its target genes. In this study we used two bioluminescent reporters to track NF-kB activation in tumor cells in response to agents that activate NF-kB. The first construct consisted of five NF-kB response elements upstream of a basal promoter controlling transcription of the luciferase gene (NF-kB-RE-luc). A second reporter was designed to link luciferase activity to lkB degradation. This construct consisted of an IkB-luciferase fusion protein driven from the EF-1alpha promoter (EF-1alpha-lkB-luc). The luciferase reporters were introduced separately and stably into human prostate carcinoma cells, PC-3M. Bioluminescence was measured in intact cells using an IVIS™ Imaging System (Xenogen Corporation). TNF-alpha and PMA increased the luciferase expression from NF-kB-RE-luc and decreased EF-1alpha-lkB-luc activity, while no effect was noted on the activity of luciferase produced from an SV-40 promoter. In the EF-1alpha-lkB-luc cell line, reduced luciferase activity correlated with a decrease in the level of IkB-luciferase fusion protein, as determined by immunoblot blot with IkB and luciferase antibodies. These cell-based assays together may be used to monitor NF-kB activation and can specifically distinguish agents that alter the stability of IkB.

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Identification of tumor associated protease substrates using combinatorial chemistry

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Prostate-specific antigen (PSA) and human kallikrein 2 (hK2) are closely related products of the human kallikrein genes KLK3 and KLK2, respectively. Both PSA and hK2 are produced and secreted by normal and malignant prostate cells. The only putative physiological substrate for hK2 include the gel-forming seminal proteins semenogelin I and II and the Nterminal propeptide activation sequence in pro PSA. Our lab is developing prodrugs and protoxins that are specifically activated by prostate tissue proteases. Although hK2 concentrations in the extracellular fluid are 100-1000 times lower than PSA, hK2 possesses \sim 1000-10,0000 fold greater enzymatic activity. Therefore, hK2 represents an attractive target for targeted prodrug therapy. To identify hK2 substrates, we synthesized a fluorescently-quenched combinatorial peptide library containing 1.6 million different peptides (one bead-one peptide). Each bead contained a unique peptide consisting of 6 randomized amino acids, flanked by the fluorophore 2-amino benzoic acid and the fluorescent guencher 3-nitrotyrosine. Incubation of this library with hK2 identified approximately 40 positive peptide sequences (beads). The determination of the peptide sequence of these digested beads is currently under investigation.